MERIT40 cooperates with BRCA2 to resolve DNA interstrand cross-links

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MERIT40 is an essential component of the RAP80 ubiquitin recognition complex that targets BRCA1 to DNA damage sites. Although this complex is required for BRCA1 foci formation, its physiologic role in DNA repair has remained enigmatic, as has its relationship to canonical DNA repair mechanisms. Surprisingly, we found that Merit40−/− mice displayed marked hypersensitivity to DNA interstrand cross-links (ICLs) but not whole-body irradiation. MERIT40 was rapidly recruited to ICL lesions prior to FANCD2, and Merit40-null cells exhibited delayed ICL unhooking coupled with reduced end resection and homologous recombination at ICL damage. Interestingly, Merit40 mutation exacerbated ICL-induced chromosome instability in the context of concomitant Brca2 deficiency but not in conjunction with Fancd2 mutation. These findings implicate MERIT40 in the earliest stages of ICL repair and define specific functional interactions between RAP80 complex-dependent ubiquitin recognition and the Fanconi anemia (FA)–BRCA ICL repair network.

[Keywords: MERIT40, BRCA2, DNA2, ubiquitination, interstrand cross-link repair]

Seminal insights into BRCA1 and BRCA2 function were revealed by their dynamic entry into discrete Rad51-containing foci in response to DNA double-strand break (DSB) formation (Scully et al. 1996, 1997; Chen et al. 1998) and essential roles in homology-directed repair (Sharan et al. 1997, Moynahan et al. 1999). This physiologic role of BRCA proteins and their interacting partners is phenotypically evident in the rare hereditary developmental disorder Fanconi anemia (FA). Germline mutations to both BRCA proteins and their interacting partners is phenotypically evident in the rare hereditary developmental disorder Fanconi anemia (FA). Germline mutations to both BRCA proteins and their interacting partners is phenotypically evident in the rare hereditary developmental disorder Fanconi anemia (FA). Germline mutations to both BRCA proteins and their interacting partners is phenotypically evident in the rare hereditary developmental disorder Fanconi anemia (FA). Germline mutations to both BRCA proteins and their interacting partners is phenotypically evident in the rare hereditary developmental disorder Fanconi anemia (FA). Germline mutations to both BRCA proteins and their interacting partners is phenotypically evident in the rare hereditary developmental disorder Fanconi anemia (FA). Germline mutations to both BRCA proteins and their interacting partners is phenotypically evident in the rare hereditary developmental disorder Fanconi anemia (FA). Germline mutations to both BRCA proteins and their interacting partners is phenotypically evident in the rare hereditary developmental disorder Fanconi anemia (FA). Germline mutations to both BRCA proteins and their interacting partners in DNA interstrand cross-link (ICL) hypersensitivity, characteristic developmental anomalies, and cancer susceptibility (D’Andrea and Grompe 2003; Deans and West 2011; Kim and D’Andrea 2012). In the case of BRCA1, a missense mutation within the BRCA1 C-terminal (BRCT) repeats disrupts interaction with ICL repair proteins BRIP1/FancJ and CtIP and with the RAP80 ubiquitin recognition complex that targets BRCA1 to DNA damage foci, resulting in a specific subtype of FA and extremely early onset breast and ovarian cancer (Domchek et al. 2013; Sawyer et al. 2015). This syndrome appears to be less severe in BRCA1 BRCT mutants than in FA patients with BRCA2 mutations. It is also noteworthy that BRCA1 BRCT mutant patients did not show bone marrow failure, which is present in patients with mutations in most other FA alleles (Domchek et al. 2013; Kottemann and Smogorzewska 2013; Garaycochea and Patel 2014; Sawyer et al. 2015).

Large BRCA foci exist as a result of DSB-induced H2AX phosphorylation (γH2AX) and ensuing recruitment of E3 ubiquitin ligases RNF8 and RNF168. Extensive RNF168-dependent ubiquitination provides a recognition platform for DNA repair proteins that span megabase stretches of chromatin surrounding DSBs (Messick and Greenberg 2009, Gudjonsson et al. 2012; Mattiroli et al. 2012; Jackson and Durocher 2013). The large BRCA1 foci require interaction with the five-member RAP80 complex [RAP80, Abraxas, MERIT40, BRCC45, and BRCC36]. Tandem RAP80 ubiquitin interaction motifs (UIMs) display specificity for non-proteasome-directed Lys63-linked ubiquitin, as does the associated deubiquitinating (DUB) enzyme BRCC36 [Sobhian et al. 2007; Sato et al. 2009; Sims and...
Cohen 2009]. RAP80 also contains several SUMO-interacting motifs just N-terminal to the UIMs and displays ~80-fold higher affinity for hybrid SUMO2,3–K63–ubiquitin chains, suggesting that both ubiquitin and SUMO chain recognition directs BRCA1–RAP80 complex foci formation (Guzzo et al. 2012). Abraxas directly binds to the BRCA1 BRCT repeats and is also required for BRCC36 DUB activity in vitro [Kim and Yu 2007; Wang et al. 2007; Feng et al. 2010; Patterson-Fortin et al. 2010]. Less well understood is MERIT40, which is not required for DUB activity in vitro but maintains interaction with the other subunits and is required for their stability in cells and localization to DNA damage foci (Feng et al. 2009; Shao et al. 2009; Wang et al. 2009). In contrast to BRCA1 nullizygosity, deficiency in either RAP80 or Abraxas is well tolerated in mice (Wu et al. 2012; Yin et al. 2012; Castillo et al. 2014), indicating that the most prominent component of DSB foci formation contributes to only a subset of BRCA1 DNA repair function (Huen et al. 2009; Jiang and Greenberg 2015).

Recent evidence suggests a physiologic role for the RAP80 ubiquitin recognition complex in ICL repair. Abraxas-null cells were sensitive to mitomycin C [MMC], and this was associated with reduced homologous recombination [HR] (Castillo et al. 2014). BRCA1 has been reported to mediate several processes during ICL repair, where it functions in a HR-independent manner to promote unloading of the replication-initiating CMG complex during early stages of ICL recognition (Long et al. 2014). BRCA1, in association with BRCA2 and Rad51, is also thought to promote HR at ICL lesions that have undergone replication-dependent processing into DSBs. RAP80 is recruited during the initial stages of ICL recognition in a ubiquitin-dependent manner, possibly implicating its function in the BRCA1-dependent CMG clearance. This step in ICL repair is important for subsequent ICL recognition by the FANCID2-I complex (Long et al. 2014).

Replication fork convergence on FANCID2-I-bound ICL lesions leads to cross-link incision and DSB formation following translesion synthesis, coupled to BRCA2–Rad51 HR repair of the resultant DSB. Evidence also exists in mammalian cells for alternative mechanisms of crosslink responses. Single-molecule imaging approaches have revealed the existence of an ICL bypass mechanism that relies on FACMC to translocate replication forks beyond psoralen cross-links (Huang et al. 2013). In addition, ICL hypersensitivity of cells deficient in the canonical D2-I pathway can be circumvented by reduced end resection [Singer 2002] increase in mortality rate was observed following treatment with the ICL-inducing agent MMC in Merit40−/− mice when compared with wild-type littermate controls [Fig. 1E]. Similarly, Merit40−/− MEFs displayed increased sensitivity to MMC, as did U2OS cells following knockdown of RAP80 complex members MERIT40 and Abraxas (Fig. 1F, Supplemental Fig. S2A). Significantly higher numbers of chromatid breaks and radial chromosome structures were also observed in Merit40−/− MEFs and primary splenocytes [Supplemental Fig. S2B, C], suggestive of an ICL repair deficit. Consistent with impaired S-phase DNA repair, apoptotic rates were elevated in Merit40−/− mice in highly proliferative organ systems 3 d after intraperitoneal injection of MMC but not in cardiomyocytes, which largely exists in a post-mitotic state [Fig. 1G,H; Supplemental Fig. S2D,E]. Together, these data highlight an important physiologic role of MERIT40 in the repair of S-phase lesions in cycling cells.

To further examine the importance of MERIT40 in S-phase DNA damage responses, wild-type and Merit40−/−
MEFs were labeled with 5-bromo-2′-deoxyuridine (BrdU) and then incubated for 1 h with MMC or hydroxyurea (HU) to halt S-phase progression. Following removal of MMC or HU, cells were chased with EdU to evaluate the ability of S-phase cells (BrdU-positive) to restart replication (Ragland et al. 2013). Merit40−/− MEFs displayed significantly reduced recovery of DNA replication in comparison with wild-type MEFs after HU or MMC treatment (Fig. 2A–C). A similar phenotype was observed when using aphidicolin (APH) to inhibit replication in both wild-type and Merit40−/− MEFs as well as in MCF10A cells with knockdown of three members of the RAP80 complex (Supplemental Fig. S3A,B). We further investigated this phenomenon using DNA fiber analysis to examine single-replication track responses in wild-type and Merit40−/− MEFs. In agreement with flow cytometry-based results, Merit40−/− MEFs displayed reduced replication restart efficiency upon transient replication blocks from MMC or HU (Fig. 2D,E). Merit40−/− MEFs were more sensitive to cisplatin, MMC, and PARP inhibitors (Supplemental Fig. S3C–E), suggestive of a broad deficiency in replication-associated DNA damage responses. Despite the significant impairment in replication restart, Merit40−/− MEFs displayed very mild sensitivity to HU, but significant elevation in chromosomal abnormalities was observed in Merit40−/− MEFs after HU treatment (Supplemental Fig. S3F,G). This pattern of mild sensitivity and increased chromosome aberrations is in agreement with prior results for Fanca mutant cells (Schlacher et al. 2012).

In addition to its role as an essential member of the RAP80 complex, MERIT40 is critical for the stability and function of the related BRISC (BRCC36 isopeptidase complex) DUB complex, which we recently implicated in promoting inflammatory cytokine receptor signaling (Cooper et al. 2009; Zheng et al. 2013). KIAA0157 is a homolog to Abraxas and is uniquely present in the BRISC. KIAA0157 knockout cells lose the BRISC while retaining.
Kiaa0157−/− MEFS showed proper recruitment of BRCA1 and RAP80 to IR-induced foci, and KIAA0157 knockout mice were not sensitive to IR or MMC (Supplemental Fig. S4). This indicates that MMC sensitivity in the absence of MERIT40 was due to deficiency in the RAP80 complex and was not related to deficiency in the BRISC.

The RAP80 complex requires PARsylation and ubiquitylation for ICL recognition

MERIT40 localizes to DSBs as a constituent of the RAP80 complex. DSB recognition requires a phosphorylation-to-ubiquitylation signaling cascade initiated by MDC1 recognition of γH2AX and subsequent recruitment of E3 ubiquitin ligases RNF8 and RNF168. To understand whether similar requirements exist for ICL repair, MERIT40 localization was monitored at UV-induced psoralen cross-links and MMC-induced cross-links (Fig. 3A, Supplemental Fig. S5A,B). The accumulation of MERIT40 at psoralen cross-links was abolished in HeLa cells upon deletion of RNF8 and RNF168 (Fig. 3A,B). Additionally, deletion of the first UIM domain eliminated the accumulation of RAP80 at psoralen cross-links (Fig. 3C,D), confirming the importance of ubiquitin binding for localization. RAP80-GFP recruitment was specific to ICL damage, as no recruitment was observed without psoralen administration prior to laser activation (Supplemental Fig. S5C). Interestingly, the initial recruitment (within 30 sec) of RAP80 was not affected by depletion of RNF8 or RNF168, indicating that rapid accumulation of RAP80 depends on other events (Supplemental Fig. S5D,E). PAR-dependent mechanisms have been implicated in the rapid localization of BRCA1 to DNA damage sites (Li and Yu 2013), and RAP80 has recently been reported to specifically bind PAR (Hu et al. 2014). Indeed, either of the PARP inhibitors NU1025 or Olaparib abrogated early phase GFP-RAP80 recruitment (<1 min) to laser-induced ICL damage (Fig. 3E,F). These data indicate that MERIT40 is recruited to ICLs in association with RAP80 in a

Figure 2. MERIT40 mediates replication fork recovery in response to S-phase damage. (A) Schematic illustration of replication fork restart experiments. Cells were pulsed with BrdU for 30 min before HU or MMC treatment to label cells that are in S phase; HU or MMC was added for 1 h to induce replication fork stalling, followed by EdU treatment for 1 h to detect fork recovery for cells in S phase. BrdU+ cells were gated, and further analysis was performed on the population to detect EdU incorporation following replication fork recovery. (B) Representative image of replication fork restart. M40 cells have a lower rate of replication fork restart. (C) Quantification of B based on three independent experiments of HU or MMC treatment. (D) Representative image of single-fiber analysis. (Red tracks) IdU; (green tracks) CldU. Example of a stalled fork and a restarted fork. (E) Schematic depicting experiment design. The percentage of restarted forks equals the number of restarted forks divided by the sum of the stalled forks and restarted forks.
biphasic manner. PARP activity is required for the initial recognition, while ubiquitin binding mediates stable accumulation of the complex.

Merit40 deficiency reduces unhooking efficiency and homology-directed repair of ICL lesions

The preceding data suggest a specific role for MERIT40 in ICL repair. As the RAP80 complex is rapidly recruited to ICL sites, we asked whether RAP80 affects FANCD2 recruitment and downstream function. RAP80 accumulated at psoralen cross-links at ~1 min, while FANCD2 required nearly 10 min for maximal ICL localization (Fig. 4A,B). Live-cell imaging experiments confirmed that mcherry-RAP80 ICL recruitment preceded that of GFP-FANCD2 (Supplemental Fig. S6A,B). FANCD2 mono-ubiquitination is essential for localization subsequent to repair of ICLs. Despite the faster ICL recruitment, knock-down of the RAP80 complex did not affect FANCD2 localization to ICL sites at 20 min after UV activation of psoralen (Supplemental Fig. S6C,D). Additionally, splenocytes from wild-type and Merit40−/− mice did not show an obvious difference with respect to FANCD2 monoubiquitination at either 4 or 24 h after MMC (Supplemental Fig. S6E).

The aforementioned results do not exclude the possibility that early recruitment of MERIT40 to ICL damage is important in the initial phases of ICL processing. A modified comet assay can be used to assess ICL unhooking efficiency in mammalian cells at the single-cell level (Hartley et al. 1999; Spanswick et al. 1999; Clauson et al. 2013). The presence of ICLs limits DNA migration into comet tails, leading to decreased tail length. The degree of ICLs present was determined by comparing the tail moment of irradiated, cisplatin-treated samples with irradiated samples that were not treated with cisplatin.
see the legend for Fig. 4; Materials and Methods for a detailed description]. The decrease in comet tail length peaked at 6 h in wild-type MEFs, followed by a sustained increase in length through 40 h. The peak in comet tail length decrease occurred at 9 h after cisplatin treatment in Merit40−/− MEFs and remained delayed in comparison with wild-type MEFs for 30 h (Fig. 4C). These data reveal that loss of MERIT40 reduces the
efficiency of ICL unhooking and support a functional importance to the rapid ICL recruitment of the RAP80 complex.

To investigate subsequent steps in ICL repair, we investigated whether loss of MERIT40 affected HR efficiency and protein recruitment. Meri40<sup>−/−</sup> cells showed modestly reduced sister chromatid exchange (SCE) after MMC treatment [Fig. 4D; Supplemental Fig. S7A]. MERIT40 knockdown was achieved by three different siRNAs in HeLa cells, and the intensity of the repair protein level was examined at psoralen cross-links. The accumulation of BRCA1 and Rad51 was reduced upon MERIT40 knockdown [Fig. 4E,F; Supplemental Fig. S7B,C]. Additionally, the mean intensity of RPA recruitment was reduced upon depletion of MERIT40 at psoralen cross-links [Fig. 4G; Supplemental Fig. S7D], and Meri40<sup>−/−</sup> cells displayed significantly reduced p-RPA levels after MMC treatment [Supplemental Fig. S7E]. Similar reductions in RPA and Rad51 at MMC-induced loci were also observed in U2OS cells following MERIT40 knockdown [Supplemental Fig. S7F,G]. These results suggest that HR deficiency in Meri40-null cells results from a cumulative reduction in end resection and Rad51 nucleofilament formation.

In contrast, several prior reports demonstrate that loss of RAP80 leads to increased end resection and HR in response to nuclease- or etoposide-induced DSBs in human cancer cell lines (Coleman and Greenberg 2011; Hu et al. 2011; Adamson et al. 2012; Goldstein and Kastan 2015). This could occur as a consequence of the RAP80 complex imparting a differential impact on the processing of DSBs in comparison with ICLs or, alternatively, due to differences between human cancer cell lines and primary mouse cells. Consistent with prior data, we detected an ~30% increase in SCEs in Meri40<sup>−/−</sup> MEFs after etoposide-induced DSBs [Supplemental Fig. 5H]. Furthermore, a 2.5-fold increase in ssDNA levels was observed in Meri40<sup>−/−</sup> splenocytes compared with wild-type counterparts, as measured by a non-denaturing BrdU signal in S/G2-phase cells at 2 h after IR [Supplemental Fig. S5I–K; Bunting et al. 2012]. These data reveal that loss of MERIT40 differentially affects end resection at DSBs and ICL damage and suggest unique contributions of the RAP80 complex to each repair process.

Meri40 deficiency does not exacerbate MMC sensitivity in Fancd2-null mice.

A genetic approach was undertaken to understand how ubiquitin recognition by the RAP80 complex intersected with the major components of ICL repair. Meri40<sup>−/−</sup> mice were bred to genetic backgrounds harboring mutations in the major arms of ICL repair, Fancd2, or conditional Brca2 (floxed exon 11). MEF cell lines were established from these crosses. Meri40, Fancd2 double-knockout mice are viable and do not show reduced body weight or other additional compromises in comparison with Fancd2-null mice. Moreover, loss of MERIT40 did not affect the sensitivity of Fancd2<sup>−/−</sup> MEFs to MMC [Fig. 5A]. Additionally, Meri40<sup>−/−</sup> Fancd2<sup>−/−</sup> MEFs or splenocytes did not show significantly different levels of chromosomal breaks or radial chromosomes in comparison with Fancd2<sup>−/−</sup> MEFs when compared with Fancd2<sup>−/−</sup> MEFS [Fig. 5B]. RPA levels at psoralen cross-links and SCEs in response to MMC were also similar in Meri40<sup>−/−</sup> Fancd2<sup>−/−</sup> MEFs when compared with Fancd2<sup>−/−</sup> MEFs [Fig. 5C–E]. However, while double-knockout Meri40<sup>−/−</sup> Fancd2<sup>−/−</sup> splenocytes did not show significant differences in total chromosomal aberrations when compared with Fancd2<sup>−/−</sup> cells, there was a trend toward increased chromosomal breaks, and a significant reduction in radial chromosomes occurred in the splenocytes of double knockouts [Supplemental Fig. S8A]. These data suggest that MERIT40 and FANCD2 are not functionally redundant during ICL repair.

Cooperative interactions between MERIT40 and BRCA2 in response to ICLs

Given the minimal genetic interaction between MERIT40 and FANC2, we asked whether MERIT40 would instead contribute in a more substantial manner to ICL repair in the context of impairment in other aspects of the FA–BRCA ICL repair network. Conditional deletion of BRCA2 was achieved in MEFs by 4-hydroxy tamoxifen (4-OHT)-induced activation of the CreERT2 fusion protein, which was constitutively expressed in cells harboring a homozygous floxed BRCA2 exon 11 allele. Wild-type and Meri40<sup>−/−</sup> MEFs expressing the CreERT2 fusion protein were used in all experiments to control for the potential off-target Cre-induced DNA damage [Silver and Livingston 2001]. Interestingly, the plating efficiency of Meri40<sup>−/−</sup> Brca2<sup>−/−</sup> MEFs was significantly lower compared with MEFs harboring either Meri40<sup>−/−</sup> or Brca2<sup>−/−</sup> in isolation [Fig. 6A,B]. Combined Brca2 and Meri40 deficiency demonstrated increased sensitivity to ICL damage, as measured by elevated cytogenetic aberrations in both MEFs and primary splenocytes [Fig. 6C; Supplemental Fig. S9A-C]. Meri40<sup>−/−</sup> Brca2<sup>−/−</sup> cells showed a greater than additive increase in chromosomal breaks compared with Meri40<sup>−/−</sup> or Brca2<sup>−/−</sup> in MEFs, while double-knockout splenocytes showed an approximately additive increase in chromosome aberrations compared with the single knockouts. Moreover, knockdown of RAP80, MERIT40, or Abraxas in pancreatic adenocarcinoma CAPAN-1 cells, which contain a truncated form of the BRCA2 protein [Goggins et al. 1996], resulted in an approximately twofold higher level of chromosomal aberrations compared with control siRNA transfected cells [Supplemental Fig. S9D]. The data are consistent with the presence of a strong genetic interaction in cells lacking MERIT40 and BRCA2 and suggest an enhanced requirement for BRCA2 in the absence of MERIT40. To understand the mechanism of how MERIT40 and BRCA2 cooperate in response to ICLs, we examined the HR efficiency by SCE analysis in response to MMC. A significantly reduced amount of SCEs was observed in Meri40<sup>−/−</sup> Brca2<sup>−/−</sup> compared with Brca2<sup>−/−</sup> cells [Fig. 6D], suggesting that MERIT40 and BRCA2 work in part to coordinately promote HR in response to ICL damage.
DNA2 differentially affects ICL repair in Merit40- and FancD2-null cells

The preceding results reveal contrasting genetic interactions between MERIT40 and canonical arms of the FA–BRCA ICL repair network: FANCD2 and BRCA2. These observations necessitated additional investigations into whether MERIT40 and ubiquitin recognition is functionally redundant with the FANCD2-I arm of ICL repair or, alternatively, constitutes a distinct component of the ICL repair network. DNA2 was previously shown to be responsible for overresection in Fancd2<sup>−/−</sup> cells, and...
knockdown of DNA2 suppressed ICL hypersensitivity in *Fancd2*−/− cells (Karanja et al. 2014). Given that ICL hypersensitivity was not increased in Merit40, *Fancd2*-double-null cells, we asked whether DNA2 knockdown would also restore ICL resistance in Merit40−/− cells. If MERIT40 affects ICL repair in a common mechanism with FANCD2, a prediction is that concomitant DNA2 deficiency would similarly impact ICL repair in either a *Fancd2*-null or *Merit40*-null background. In agreement with previous reports (Karanja et al. 2014), DNA2 knockdown reduced genomic instability in MMC-treated *Fancd2*−/− cells and reduced cell survival in response to MMC (Fig. 6E; Supplemental Figs. S8D, S10A). These data reveal that MERIT40’s and FANCD2’s contributions to ICL repair can be differentiated by their genetic interactions with DNA2.

DNA2 knockdown also increased genomic instability in *Merit40−/− Brca2−/−* cells (Fig. 6E, Supplemental Fig. S10A). Similar results were obtained following transfection of siRNA targeting CtIP in double-knockout cells and by Exo1 knockdown in *Merit40−/−* MEFS, consistent with an increased requirement for end resection in MERIT40-null cells (Supplemental Fig. S10B,C). Comparable SCE numbers were observed before and after DNA2 knockdown in *Merit40−/− Brca2−/−* cells (Fig. 6F), suggesting that DNA2-dependent resection contributes to an alternative, HR-independent repair function in the context of dual deficiency for BRCA2 and MERIT40. Knockdown of essential microhomology-mediated end-joining

**Figure 6.** Cooperative genetic interactions occur between MERIT40 and BRCA2 in response to ICL damage. (A,B) Plating efficiency of wild type (WT), M40, two clones of inducible *BRCA2*Δ/Δ*Cre-ERT2* (B2A and B2B), and three clones of inducible MERIT40 *BRCA2*Δ/Δ*Cre-ERT2* MEFS (M40B2A, M40B2B, and M40B2C). MEFS were treated with 500 nM 4-OHT for 48 h and then plated at 500 cells per 60-mm plate. Crystal violet-stained colonies were counted 10 d after plating. Student’s t-tests were performed between wild type and M40A and also between B2A and three clones of M40 B2 MEFS. (C) Wild-type, M40, B2/Δ*Cre-ERT2*, and M40 B2/Δ*Cre-ERT2* MEFS were treated with 4-OHT for 48 h to induce *BRCA2* incision. Cells were then treated with 20 ng/mL MMC for an additional 24 h prior to metaphase harvest. *n* > 100 metaphases per genotype. Student’s t-tests were performed between B2 and M40 B2 cells. (D) MEFS were incubated with 4-OHT for 48 h to induce *BRCA2* incision and then treated as in Figure 4D. Student’s t-tests were performed between B2 and M40 B2 cells, and quantification was derived from four independent experiments. (E) Loss of DNA2 exacerbates levels of chromosomal abnormalities in *M40 B2* MEFS. Two independent experiments were conducted with two different siRNAs. The data for siDNA2-1 are shown in Supplemental Figure 10A. (F) SCEs were analyzed for the indicated cells. Student’s t-tests were performed between M40 B2 siCtrl and M40 B2 siDNA2-2. Two independent experiments were conducted.
(MMEs) proteins Lig3 or POLQ did not exacerbate chromosomal abnormalities (Supplemental Fig. S1D–F), suggesting that other resection-dependent repair events are invoked in Meri40−/− Brca2−/− cells.

Discussion

The physiologic importance of RAP80 complex ubiquitin foci association has remained elusive despite considerable investigation into DNA damage recognition mechanisms and its roles in cancer susceptibility (Nikkilä et al. 2009; Antoniou et al. 2010; Bolton et al. 2010; Solyom et al. 2012; Castillo et al. 2014). This study reveals that a major function of the RAP80 complex is in DNA replication-associated repair. Surprisingly, in contrast to Rap80−/− and Abraxas−/− mice (Wu et al. 2012; Yin et al. 2012; Castillo et al. 2014). Meri40−/− mice or cells were not significantly hypersensitive to IR even though they exhibited G2 checkpoint impairment. These issues notwithstanding, Abraxas deficiency was also reported to confer MMC hypersensitivity (Castillo et al. 2014). Our results are consistent with these published reports in that deficiencies in multiple RAP80 complex members showed similar deficits in ICL responses.

ICL hypersensitivity was associated with reduced ssDNA generation and homology-directed repair in Meri40−/− cells despite elevation of these DNA damage-associated markers in response to etoposide-IR-, or nuclease-induced breaks (Coleman and Greenberg 2011; Hu et al. 2011; Goldstein and Kastan 2015). The opposing reduction in ssDNA formation in response to ICL damage may be a consequence of delayed ICL unhooking in Meri40−/− cells, consistent with involvement of MERI40 in the earliest stages of ICL recognition and processing. Several nucleases have been suggested to process ICL lesions, including the SLX4 complex and FAN1 (Kratz et al. 2010; MacKay et al. 2010; Smogorzewska et al. 2010; Tsai et al. 2010; Crossan et al. 2011; Kim et al. 2011; Stoepker et al. 2011; Kim and D’Andrea 2012; Kottemann and Smogorzewska 2013). Future investigation is warranted as to whether MERI40 affects their activities at ICLs.

Meri40−/− MEFs also displayed difficulty in restarting replication forks after DNA polymerase stalling (Fig. 2; Supplemental Fig. S3A,B), indicating that, in addition to ICL repair, the primary role of MERI40 function is more broadly applicable to S-phase genotoxic stress. Multiple FA and BRCA pathway proteins have been implicated in replication fork maintenance or restart (Petermann et al. 2010; Schlacher et al. 2011, 2012; Neelsen and Lopes 2015; Zellweger et al. 2015). The function that MERI40 exerts in S phase might be through its direct role in replication fork restart or through recruitment of BRCA1, which has been established to protect replication forks from nucleases (Schlacher et al. 2012). DNA2 has also been shown to mediate fork restart after MMC treatment by degrading reversed replication forks and preventing the formation of aberrant replication fork intermediates (Thangavel et al. 2015). The fact that knockdown of DNA2 in Meri40−/− MEFs led to lower plating efficiency without any genotoxic treatment (data not shown) indicates that MERI40 and DNA2 might work in independent processes to maintain replication fork stability during ICL repair.

Repair of ICL lesions in S phase requires a multistep process that entails ICL recognition, dual incisions on either side of the cross-link, DNA synthesis through the cross-link by TLS polymerases and BRCA–Rad51-dependent HR repair of the ensuing DSB. The ubiquitinated D2-I complex plays a critical role in early ICL recognition and processing steps, whereas BRCA1 and BRCA2 have been implicated in both early and later aspects of ICL repair. Our extensive genetic interaction data are supportive of epistasis between MERI40 and FANCD2. MERI40 was required for timely ICL unhooking, and, importantly, double-knockout Meri40−/−, FancD2−/− mice and cells did not show any phenotypic or ICL repair exacerbations beyond that of FancD2 alone. In stark contrast, Meri40−/− and Brca2−/− alleles produced at least additive increases in genomic instability in response to MMC coupled with strongly reduced cell viability. However, we posit that MERI40 contributes to multiple aspects of ICL repair (Fig. 7) in a manner that is not completely overlapping with FANCD2-1, thus accounting for differences in its genetic interactions with DNA2. These genetic interactions enhance our understanding of the multifactorial
response to ICL damage and suggest targeting MERIT40 or other components of the RAP80 complex as an additional means to reduce viability in BRCA2 mutant cells.

Materials and methods

Mice

C57BL/6J [strain 000664] and B6(Cg-TyrC-C2)]/J [strain 00058] were purchased from the Jackson Laboratory. The homozygous conditional BRCA2<sup>mut</sup> mouse strain [01XB9] was purchased from the National Cancer Institute Mouse Repository. The CreERT2 B6.Cg-Tg(UBC-cre/ERT2)1EjB/J strain was generously provided by Dr. Eric J. Brown [University of Pennsylvania, Philadelphia]. The FancD2 knockout mouse was generously provided by Dr. Alan D’Andrea [Dana-Farber Cancer Institute, Harvard Medical School].

Generation of MERIT40 knockout mice, genotyping, and MEFs

MERIT40 (RIKEN cDNA 5430437P03 gene MGI: 1915501) knockout mice were generated by microinjecting an embryonic stem cell clone provided by Texas A&M Institute for Genomic Medicine [IST11207B9] into C57BL/6J host mouse blastocysts. The MERIT40 locus was disrupted by insertion of the Omnibank expression vector 76 into the MERIT40 first intron upstream of the MERIT40 coding sequence. Generation of MERIT40 knockout mice, genotyping, and MEFs were carried out as described in this section.

Genotyping

Genotyping was performed by PCR on genomic DNA isolated from mouse tails by digestion overnight at 55°C in a lysis buffer containing 100 mM Tris [pH 8.0], 5 mM EDTA, 0.2% SDS, 200 mM NaCl, and 100 µg/ml proteinase K. DNA was precipitated with isopropanol and washed twice with 70% ethanol before being resuspended in TE buffer, quantified, and used for PCR reactions. The following primers were used for genotyping: M40F (5′-GGGAAAACCTCACTTATCTTTCCGCTC3′), M40R (5′-AGGACGCAATGATGCCGACAGCCCCCTTAC3′), and V76R (5′-CTTGCAAAAATGCGTATAAGCC-3′). Primer pair M40F/M40R gave a band of 480 base pairs [bp] corresponding to the wild-type allele, and PCR from primer pair M40F/V76R gave a band of 320 bp corresponding to the disrupted allele. PCR conditions did not allow formation of the product corresponding to the wild-type allele when containing the gene trap vector. Amplification reactions were performed using GoTaq Green mastermix (Promega) in a S1000 thermal cycler (Bio-Rad). BRCA1 and FancD2 matched (between 6 and 12 wk) wild-type and MERIT40<sup>−/−</sup> mice. To assess radiosensitivity and MMC sensitivity in vivo, age-matched [between 6 and 12 wk] wild-type and Merit40<sup>−/−</sup> littermates were subjected to 8.76 Gy of IR or 12 mg/kg MMC. MMC was resuspended in PBS and injected intraperitoneally. Mice were monitored daily to establish Kaplan-Meier curves. A log-rank test was used for statistical analysis.

Clonogenic survival assay

Cells were induced with 500 nM 4-OHT for 48 h and then seeded in a 60-mm dish. The following day, cells were treated with MMC at the indicated dose for 24 h. The drug was washed out, and cells were allowed to recover for 1–2 wk. Plates were stained with 0.5% crystal violet for 1 h, and colonies were counted. Each genotype and drug dose was done in triplicate in three independent experiments.

Cell culture

Mouse survival assay

To assess radiosensitivity and MMC sensitivity in vivo, age-matched [between 6 and 12 wk] wild-type and Merit40<sup>−/−</sup> littermates were subjected to 8.76 Gy of IR or 12 mg/kg MMC. Rap80<sup>−/−</sup> and Rap80<sup>−/−</sup> cell lines were provided by Dr. Junjie Chen [M.D. Anderson Cancer Center]. MEFs were immortalized after transfection with an shRNA directed against p19Arf. Spleenocytes were maintained in RPMI glutamax with β-mercaptoethanol, 10% FBS, 1% sodium pyruvate, 1% nonessential amino acid, and 10 µg/mL LPS.

Antibodies

A MERIT40 rabbit polyclonal antibody raised against mouse MERIT40 was generated against a GST-MERIT40 fusion protein and used at 1:200 for immunoblotting and 1:100 for immunofluorescence as previously described (Shao et al. 2009). γ-H2AX (Millipore, JBW301) was used at 1:2000 for immunofluorescence and 1:1000 for immunoblotting. BRCA1 was detected by immunofluorescence with a homemade rabbit polyclonal antibody raised against the exon 11 region of mouse BRCA1 at 1:100. Homemade Rap80 rabbit polyclonal antibody was used for immunoblotting at 1:500 and immunofluorescence at 1:150. BRCC36 was detected for immunoblotting with a rabbit polyclonal antibody (LSBio, LS-C102632) diluted at 1:1000 [Zheng et al. 2013]. α-Tubulin [1:10,000, Cell Signaling,
DM1A was used as a loading control on immunoblots. p-RPA (S4/S8) [Bethyl Laboratories, A300-245a] was used for immunoblotting at 1:1000. Total RPA (Novus Biologicals, NB600-565) was used for immunoblotting at 1:200. FANCID2 (Abcam, ab108928) was used of immunoblotting at 1:200. RPA32/RPA2 (Abcam, ab61184) was used for immunofluorescence in mouse cells at 1:200.

Immunofluorescence

Cells were washed with PBS and pre-extracted in a buffer containing 10 mM Pipes, 100 mM NaCl, 300 mM sucrose, 3 mM MgCl2, and 0.5% Triton X-100 for 5 min at 4°C or fixed directly in a solution containing 3% paraformaldehyde and 2% sucrose for 10 min at room temperature. Cells were subsequently permeabilized with 0.5% Triton-X100, 50 mM Tris-HCl (pH 7.4), 150 mM NaCl, 5 mM EDTA, 0.05% NP-40, and 0.25% gelatin IV for 10 min at 4°C and then incubated in a blocking solution (10% goat serum in PBST) prior to incubation with the appropriate primary antibody for 1 h at 37°C. Cells were then washed with PBST and next incubated with secondary antibody diluted in PBST for 20 min at 37°C. After three washes with PBST, coverslips were mounted onto glass slides using Vectashield mounting medium containing DAPI (Vector Laboratories) and visualized using a Nikon Eclipse 80i fluorescent microscope.

Metaphase spreads

Cells were treated with 0.5 μM nocodazole for 3 h and swollen in 75 mM KCl for 25 min at 37°C. Cells were fixed on ice with a 3:1 methanol/acetic acid solution. Metaphases were dropped onto slides preheated at 42°C, allowed to dry, and stained with Giemsa. The numbers of sister chromatid breaks and chromosome breaks per metaphase were counted. For SCE analysis, cells were pretreated for 48 h with 10 μM BrdU and collected as above. After metaphases were dropped onto slides, they were stained with 10 μg/mL Hoechst 33258 in PBS for 20 min. The slides were rinsed in McIlvaine solution for 10 min and treated with 365-nm UV for 30 min. Slides were incubated in 1× SSC for 1 h at 55°C and then stained with Giemsa and analyzed for the number of SCEs per chromosome.

DNA fiber assay

Cells were labeled with 50 μM IdU or 100 μM CldU. DNA fibers were spread as described [Jackson and Pombo 1998] and stained with primary and fluorescent secondary antibody [primary antibodies: α-IdU [Rector Dickinson, 347580] and α-CldU [Abcam, ab6326]]. Fibers were imaged with a 60x objective and analyzed using ImageJ software.

Native BrdU detection

Exponentially growing splenocytes were pulsed with 1 mM BrdU (Sigma) for 30 h, irradiated (30 Gy; 2 h of recovery at 37°C), and then fixed with 70% ethanol. Cells were blocked in 3% BSA in PBS for 30 min and then stained with anti-BrdU monoclonal antibody [1:50, Invitrogen, Mobu-1] for 1 h. Before analysis, propidium iodide was added to a final concentration of 50 μg/mL.

Modified alkaline comet assay

Cells were treated with 50 μM cisplatin for 1 h. After treatment, cells were either immediately harvested (0 h) or incubated in fresh medium before harvesting at 4, 6, 7.5, 9, 17, or 40 h after cisplatin was washed off. Control cells were not treated with cisplatin. Immediately before analysis, cells were irradiated (12.5 Gy) to get a fixed number of DNA strand breaks. We performed an alkaline comet assay based on the protocol from CometAssay kit [Trevigen, 42150-050-K]. Olive tail moment was obtained by using the OpenComet plug-in in ImageJ. The percent decrease in Olive tail moment = [1 – (TMdi – TMcu)/(TMci – TMcu)] × 100, where TMdi is the tail moment of the cisplatin-treated, irradiated sample; TMcu is the tail moment of the untreated, unirradiated control; and TMci is the tail moment of the untreated, irradiated control. The percent decrease in Olive tail moment at the early time point demonstrates peaks when ICLs in cells get unhooked.

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References


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References


**MERIT40 is essential for ICL repair**


MERIT40 cooperates with BRCA2 to resolve DNA interstrand cross-links

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