Integrin-linked kinase, a novel component of the cardiac mechanical stretch sensor, controls contractility in the zebrafish heart

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The vertebrate heart possesses autoregulatory mechanisms enabling it first to sense and then to adapt its force of contraction to continually changing demands. The molecular components of the cardiac mechanical stretch sensor are mostly unknown but of immense medical importance, since dysfunction of this sensing machinery is suspected to be responsible for a significant proportion of human heart failure. In the hearts of the ethynitros-urea (ENU)-induced, recessive embryonic lethal zebrafish heart failure mutant main squeeze (msq), we find stretch-responsive genes such as atrial natriuretic factor (anf) and vascular endothelial growth factor (vegf) severely down-regulated. We demonstrate through positional cloning that heart failure in msq mutants is due to a mutation in the integrin-linked kinase (ilk) gene. ILK specifically localizes to costameres and sarcomeric Z-discs. The msq mutation (L308P) reduces ILK kinase activity and disrupts binding of ILK to the Z-disc adaptor protein β-parvin (Affixin). Accordingly, in msq mutant embryos, heart failure can be suppressed by expression of ILK, and also of a constitutively active form of Protein Kinase B (PKB), and VEGF. Furthermore, antisense-mediated abrogation of zebrafish β-parvin phenocopies the msq phenotype. Thus, we provide evidence that the heart uses the Integrin–ILK–β-parvin network to sense mechanical stretch and respond with increased expression of ANF and VEGF, the latter of which was recently shown to augment cardiac force by increasing the heart’s calcium transients.

[Keywords: Integrin-linked kinase [ILK], zebrafish, cardiac stretch sensor, β-parvin [Affixin]]

Supplemental material is available at http://www.genesdev.org.

Received May 9, 2006; revised version accepted July 7, 2006.

The vertebrate heart possesses a unique autoregulatory mechanism enabling it to adapt its force of contraction to continually changing demands. On a beat-to-beat basis, the heart can adapt to changes in blood pressure or volume load. Chronic overload induces long-term changes that augment contractile force by assembly of additional myofibrils, with consequent cardiac hypertrophy. These adaptive responses of the heart are preceded by the sensing of biomechanical strain (mechanosensing) and transduction of these signals (mechanotransduction). Mechanical stretch of cardiomyocytes activates several signaling pathways, including mitogen-activated protein [MAP] kinases, JAK–STAT, and phosphatidylinositol 3-kinase [PI3K]–AKT [Pan et al. 1999, Seko et al. 1999, Kim et al. 2002], ultimately inducing the expression of various cardiac stretch-responsive genes, such as atrial natriuretic factor (anf) and vascular endothelial growth factor (vegf) [Kim et al. 2002]. However, most of the molecular components of the mechanical stretch sensor that trigger these cascades in the heart are still unknown. They are additionally of great clinical importance, because these mechanisms are part of the adaptive response to cardiac disease and of heart failure. In fact, dysfunction of components of the cardiac stress sensor system, including β-integrins, the integrin-interacting protein Melusin, or the Z-disc protein Muscle lim protein (MLP), can cause heart failure [Shai et al. 2002].

Integrin-linked kinase [ILK], an evolutionarily highly conserved serine/threonine [Ser/Thr] protein kinase, is expressed at high levels in cardiac and skeletal muscle, where it interacts with the cytoplasmic domain of β1-Integrin [Hannigan et al. 1996]. ILK contains three identifiable structural features, an ankyrin repeat region at
ILKBP, Actopaxin), lack of ILK expression in the G-actin sequestering peptide Thymosin-4 intercalates with Protein Kinase B (PKB). The adaptor protein PINCH and protein kinase domain: the transmembrane receptor protein, at least five different proteins bind to the C-terminally located kinase domain: the transmembrane receptor protein-β1-Integrin, the adaptor proteins α-parvin (CH-ILKBP, Actopaxin), β-parvin (Affixin), Paxillin, and the catalytic protein PKB. The adaptor protein PINCH and the G-actin sequestering peptide Thymosin-β4 interconnect with the N-terminal ankyrin repeat domain of ILK (Bock-Marquette et al. 2004). Lack of ILK expression in Drosophila melanogaster and Caenorhabditis elegans results in severe muscle abnormalities (Zervas et al. 2001; Mackinnon et al. 2002), whereas mice lacking ILK expression die shortly after implantation, indicating an essential role of ILK in early embryonic development of vertebrates. However, the structural and functional roles of ILK in the vertebrate heart are still unknown.

To identify novel molecular components of the cardiac stretch sensor, we have assayed here the cardiac expression of anf and vegf in recessive embryonic lethal zebrafish heart failure mutants (T. Dahme, I.G. Huttner, and W. Rottbauer, unpubl.). Both anf and vegf were found to be significantly down-regulated in the hearts of the zebrafish mutant main squeeze (msq). By positional cloning we show that msq mutants carry a mutation in the zebrafish integrin-linked kinase (zilk) gene, which specifically disrupts ILK kinase activity and ILK-β-parvin binding. Heart contractility of homozygous mutant msq embryos can be restored by ectopic expression of constitutively active PKB or wild-type VEGF, and morpholino-modified antisense oligonucleotide mediated knockdown of zebrafish β-parvin phenocopies the msq heart phenotype. Taken together, our findings indicate that ILK, via interaction with Integrin and β-parvin, acts as an essential component of the cardiac stretch sensor to control transcription of the stretch-responsive genes anf and vegf and cardiac contractility.

Results

Stretch-responsive genes anf and vegf are severely down-regulated in zebrafish main squeeze (msq) hearts

To identify novel genetic components of the cardiac stretch sensor, we screened numerous ethylnitros-urea (ENU)-induced recessive, embryonic lethal zebrafish heart failure mutants [Stainier et al. 1996] for altered cardiac expression of the stretch-responsive genes anf and vegf, using whole-mount RNA antisense in situ hybridization and real-time PCR (T. Dahme, I.G. Huttner, and W. Rottbauer, unpubl.). In the hearts of the zebrafish mutant main squeeze (msqm347), we found both anf [Fig. 1C–F] and vegf severely down-regulated at 72 h post-fertilization (hpf).

msq mutant embryos display progressive reduction of cardiac contractility. Aside from pericardial edema, msq embryos are not noticeably affected by the lack of normal blood flow during the first week of development [Fig. 1A,B]. As in wild-type embryos, the two heart chambers—atrium and ventricle—contract rhythmically, sequentially, and vigorously in msq mutant embryos by 36 hpf. However, by 72 hpf, ventricular contractility of msq mutant embryos decreases progressively: fractional shortening of the msq ventricular chamber declines from 20.9 ± 4.9% at 48 hpf to 1.5 ± 3.26% at 72 hpf, and by 96 hpf, the ventricular chamber becomes completely silent [Fig. 1G,H; Supplementary Movie 1].

To determine whether the msq heart contractility defect is due to abnormal growth of ventricular cardiomyocytes as observed for the zebrafish mutants island beat [Rottbauer et al. 2001] and liebeskummer [Rottbauer et al. 2002], or due to disturbed cardiomyocyte myofibrillogenesis, as observed in the mutants pickwik [Xu et al. 2002], silent heart [Sehnert et al. 2002], or tell tale heart [Rottbauer et al. 2006], we analyzed the msq mutant hearts structurally and ultrastructurally. As in the wild type, embryonic heart morphogenesis proceeds normally in msq mutant embryos. By 72 hpf, msq mutant ventricles are structurally indistinguishable from wild-type hearts: A myocardial as well as an endocardial layer is clearly present, and the ventricular myocardium has thickened properly by addition of myocardial cells. The number of msq ventricular cardiomyocytes [330 ± 10 SEM] is indistinguishable from wild type [340 ± 22 SEM] [Fig. 1J], and there is no increase in the number of apoptotic myocardial cells compared with wild-type embryos assayed by TUNEL staining [data not shown]. By whole-mount immunofluorescence, we investigated the expression of thick and thin filament sarcomeric proteins in wild-type and msq mutant embryos. Sarcomeric atrial and ventricular myosin heavy chain as well as sarcomeric actin protein expression is normal in msq mutant hearts [data not shown]. Transmission electron microscopy of msq hearts after 72 hpf reveals a normal architecture of ventricular cardiomyocytes, including regular arrays of thick and thin myofilaments, Z-discs, and intercalated discs, as well as normal content and appearance of mitochondria [Fig. 1K,L]. These findings indicate that msq does not disrupt key steps in zebrafish heart morphogenesis or myocardial myofibrillogenesis, but rather interferes with an as-yet-unknown essential functional component of the contractile machinery.

The main squeeze (msqm347) locus encodes zilk

We identified the ENU-induced mutation causing the recessive msq mutant phenotype by a positional walk [Fig. 2A]. By bulk segregant analysis, we mapped msq to zebrafish linkage group 10. Recombination analysis of
1674 msq mutant embryos restricted msq to a genomic interval flanked by the microsatellite markers z7504 and z7028 (Fig. 2A). According to genomic sequence alignment (BLASTX), the physical contig covering the msq locus contains three ORFs encoding proteins highly homologous to human ILK, Sphingomyelinase, and an as-yet-unknown protein (GenBank accession no. Riken 2210415M20).

To identify the site of the ENU-induced mutation in msq, we sequenced the entire zebrafish coding sequences of the three ORFs from wild-type and msq mutant cDNA and gDNA. Sequencing of neither the zebrafish sphingomyelinase nor Riken 2210415M20 coding sequence revealed a mutation in msq mutant cDNA. However, we identified the msq mutation to be a thymine-to-cytosine nucleotide transversion (CTA → CCA) in codon 308 of the zilk gene, predicted to change the highly conserved amino acid leucine to proline (L308P) (Fig. 2B). zilk is endcoded by 11 exons. The protein consists of 452 amino acids and shows 86% amino acid identity to human and mouse ILK. As in other species, zILK contains at its N-terminal four ankyrin (ANK) repeats, followed by a PH domain and a kinase catalytic domain (amino acids 192–420), where the msq mutation (L308P) resides (Fig. 2C).

Injection of morpholino-modified antisense oligonucleotides against zilk phenocopies the msq heart phenotype

To confirm that the msq mutant heart phenotype is caused by disturbed zILK function, we injected morpholino-modified antisense oligonucleotides, either directed against the translational start site [MO1-zilk] or the splice donor site of exon 4 [MO2-zilk] of zilk into one-cell-stage wild-type zebrafish embryos. When injecting either 4.2 ng of MO1-zilk or 2.5 ng of MO2-zilk, up to 87% of injected embryos (39 out of 68 for MO1-zilk and 85 out of 119 for MO2-zilk) display the msq mutant heart phenotype [Fig. 3A–C]. Similar to msq mutant embryos, in MO1-zilk- and MO2-zilk-injected embryos the
expression of the stretch-responsive gene *zanf* is severely down-regulated (Fig. 3D–G), and fractional shortening of the ventricular chamber consecutively decreases to 0.63 ± 1.77% at 72 hpf (Fig. 3H; Supplementary Movie 2).

Isolation of RNA from MO2-*zilk*–injected embryos confirmed the predicted effect on *zilk* mRNA splicing, namely, either skipping of exon 4 (125-base-pair [bp] product) or integration of intron 4 (370-bp product), both leading to premature termination of translation (Fig. 3H, inset). Wild-type *zilk* cDNA (224-bp product) was not detected in the injected embryos even 72 h after injection. These findings implicate that loss of cardiac ventricular contractility and reduced expression of the stretch-responsive genes *anf* and *vegf* in *msq* mutants are due to loss of ILK function.

**ilk mRNA restores ventricular contractility in msq mutant embryos**

Our morpholino knockdown experiments imply that the *msq* mutation interferes with an essential function of *zILK* in ventricular cardiomyocytes. We therefore evaluated whether ventricular contractility in *msq* mutant embryos can be restored by ectopic expression of wild-type *zilk* mRNA. After injection of 300 pg of wild-type *zilk* mRNA (*zilk*wt), 83% of the injected *msq* mutant embryos (24 out of 29) displayed normal ventricular function by 72 hpf. In contrast, injection of *msq* mutant *zilk* mRNA (*zilk*L308P) at various concentrations (300–500 pg) did not rescue the contractile dysfunction in *msq* mutant embryos (0 out of 50) (Fig. 3I).

As described above, *zILK* and *hILK* show 86% amino acid identity (Fig. 2C). To test if the functional role of ILK in maintaining cardiac ventricular contractility is conserved between zebrafish and human, we injected human *ilk* mRNA (*hilk*wt) into one-cell-stage embryos derived from intercrossing *msq* heterozygous +/− zebrafish. When injecting 500 pg of full-length *hilk*wt, 43% of *msq* mutant embryos (18 out of 42) displayed normal ventricular contractility at 72 hpf (Fig. 3I).
Ectopic expression of wild-type ILK results in products of 380 bp (integration of intron 4) and 125 bp (skipping of exon 4), predicted to lead to premature termination of translation of zmsq mRNA expression in the heart and skeletal muscle, blood vessels, and the pancreas (Hannigan et al. 1996; Zervas et al. 2001; Mackinnon et al. 2002). Similarly, by 72 hpf, expression is pronounced in the heart and skeletal muscle, blood vessels, and the pancreas (Hannigan et al. 1996; Zervas et al. 2001; Mackinnon et al. 2002). Similarly, by 72 hpf, expression is pronounced in the heart and skeletal muscle, blood vessels, and the pancreas (Hannigan et al. 1996; Zervas et al. 2001; Mackinnon et al. 2002).

These data confirm that the zILK L308P mutation is, indeed, responsible for the msq cardiac phenotype, and imply that the function of ILK in maintaining cardiac contractility is conserved between zebrafish and human.

zILK specifically localizes to costameres and Z-discs of heart and skeletal muscle

In other species, Ilk is widely expressed both during embryonic development and in the adult, with highest expression observed in heart and skeletal muscle, blood vessels, and the pancreas (Hannigan et al. 1996; Zervas et al. 2001; Mackinnon et al. 2002). Similarly, by 72 hpf, zilk mRNA expression is pronounced in the heart and skeletal muscle of zebrafish embryos. zilk mRNA expression levels do not differ between wild-type and msq mutant embryos (data not shown).

To further determine the localization of zILK protein in the heart and within cardiomyocytes, we performed immunostaining with a monoclonal antibody raised against full-length human ILK [Li et al. 1999]. As shown in Figure 4A, ILK expression can be detected both in the endocardial and myocardial layer of the zebrafish embryonic heart. Within cardiomyocytes, zILK shows a striated distribution and colocalizes with the sarcomeric Z-disc protein α-Actinin [Fig. 4B,C]. A similar subcellular localization can be observed in skeletal muscle cells [Fig. 4D], where zILK also localizes to the sarcolemma, enriched at those areas of the cell membrane where Z-bands anchor. The same pattern is observed in adult zebrafish [Fig. 4E] and rat cardiomyocytes [Fig. 4F]. In addition, immunogold staining and consecutive transmission electron microscopy confirms that some ILK protein localizes directly to the sarcomeric Z-disc of cardiomyocytes [Fig. 4G]. Taken together, these findings demonstrate that in cardiomyocytes and skeletal muscle cells, ILK is concentrated to regions on the sarcolemma, where Z-bands attach, the so-called costameres.

To further evaluate if the msq mutation ILK L308P interferes with zILK stability and its subcellular localization to costameres and Z-discs in cardiomyocytes and skeletal muscle cells, we analyzed zILK expression in msq mutant zebrafish embryos by immunostaining. As shown in Figure 4, H and I, the ILK antibody readily recognizes mutant ILK L308P, and there is no difference in the expression level and intracellular localization of zILK in comparison to wild-type zebrafish embryos [Fig. 4A,D], indicating that msq mutant ILK L308P protein is stable and localizes correctly to costameric units of heart and skeletal muscle cells.

The msq ILK L308P mutation leads to reduced ILK kinase activity

The msq mutation ILK L308P resides within the highly conserved serine/threonine kinase domain of ILK [Delcommenne et al. 1998; Yamaji et al. 2001]. Specific residues within the kinase domain, such as the arginine residue at position 211, are known to be essential for ILK kinase activity [Persad et al. 2001a].
ILK
tant ILK308P is expressed at normal levels in cardiac muscle and sarcomeric Z-discs. ([A] ILK is expressed in the endocardial [en] and myocardial [my] layer of the zebrafish heart. Cross-section through the cardiac ventricle of a wild-type zebrafish embryo at 72 hpf. [B–C] Coimmunostaining with an antibody directed against the sarcomeric Z-disc protein α-Actinin reveals colocalization of ILK and α-Actinin in zebrafish cardiac myocytes. [C] Merge of the anti-ILK and anti-α-Actinin stain. [D] As already observed in cardiomyocytes (C), ILK and α-Actinin also colocalize at sarcomeric Z-discs of zebrafish skeletal muscle cells. In addition, as indicated by arrowheads, sarcolemmal localization of ILK can be observed. ([E–G] In the adult zebrafish [E] and rat [F,G] heart, ILK and α-Actinin also colocalize at sarcomeric Z-discs. [G] Immuno-gold staining and transmission electron microscopy of rat heart sections confirms localization of ILK at sarcomeric Z-discs. [H] msq mutant ILK<sup>L308P</sup> is expressed at normal levels in msq mutant zebrafish hearts [H] and skeletal muscle [I], and shows sarcolemmal staining (indicated by arrowheads) and colocalization with α-Actinin.

Therefore, to examine if the msq mutation impairs ILK kinase activity and thereby reduces cardiac contractility, we assayed the kinase activity of msq mutant [zILK<sup>L308P</sup>] and wild-type [zILK<sup>wt</sup>] ILK in vitro. As shown in Figure 5A, phosphorylation of MBP by msq mutant zILK<sup>L308P</sup> was reduced by ∼58% in comparison to phosphorylation by wild-type zILK<sup>wt</sup>. Similarly, autophosphorylation of zILK<sup>L308P</sup> was reduced by ∼50% in comparison to wild-type zILK<sup>wt</sup> [Fig. 5A]. To further evaluate whether reduction of zILK<sup>L308P</sup> kinase activity accounts for the msq mutant heart phenotype in vivo, we evaluated the ability of kinase-deficient human ILK [hILK<sup>R231A</sup>] (Persad et al. 2001a) to reconstitute cardiac contractility in msq mutants. Strikingly, in contrast to hILK<sup>R231A</sup> mRNA, injection of even high amounts (300–800 pg) of kinase-deficient hILK<sup>R231A</sup> mRNA was not sufficient to suppress the cardiac phenotype in homozygous mutant msq embryos (0 out of 71) [Fig. 5B]. In summary, our findings indicate that reduced kinase activity of msq mutant zILK<sup>L308P</sup> might account for loss of cardiac contractility in msq mutant embryos.

PKB and VEGF act downstream of ILK in controlling vertebrate heart contractility
ILK directly phosphorylates PKB at Ser 473, thereby leading to full activation of PKB [Persad et al. 2001a]. Furthermore, ILK controls transcription of vegf via PKB phosphorylation [Tan et al. 2004]. We recently demonstrated that VEGF controls calcium cycling and contractility of the zebrafish embryonic heart [Rottbauer et al. 2005]. Therefore, we speculated that reduced kinase activity of msq mutant ILK might disturb ILK–PKB–VEGF signaling in cardiomyocytes, leading to the msq mutant heart phenotype.

To further elucidate whether PKB acts downstream of ILK to control cardiac contractility in msq mutant hearts, we injected constitutively active, myristoylated PKB [myrPKB] mRNA, which was recently shown to be functional in zebrafish [Chan et al. 2002] into msq mutant embryos. After injection of 500 pg of myrPKB mRNA, 53% of the injected msq mutant embryos (36 out of 68) displayed normal ventricular contractility by 72 hpf [Fig. 5B], whereas injection of a kinase-dead version of myristoylated PKB [inactPKB] mRNA at various concentrations (500–800 pg) had no effect on the contractile dysfunction in msq mutant embryos [0 out of 68] [Fig. 5B], indicating that reduced PKB activity downstream of ILK might account for heart failure of msq mutants. Interestingly, in whole-embryo lysates, phosphorylation of PKB at Ser 473 was not significantly different between wild-type and msq mutants, indicating that besides ILK other kinases might contribute to full activation of PKB, at least in tissues other than the heart.

In our initial screen, VEGF expression was found to be sixfold down-regulated in msq mutant hearts. To test whether reduced VEGF expression is directly involved in loss of cardiac contractility in msq mutants, we injected zvegf<sub>121</sub> mRNA into msq mutant embryos and evaluated their heart function. After injection of 250 pg of zvegf<sub>121</sub> mRNA, 10% of the injected msq mutant embryos (3 out of 30) displayed completely restored ventricular function by 84 hpf [Fig. 5B]. Additional 17% (5 out of 30) of injected msq mutant embryos showed weak ventricular
contractions, resembling partial rescue of msq mutant heart failure. These findings indicate that VEGF can restore ventricular contractility in msq mutant embryos and might act downstream of ILK and PKB in the cardiac stretch sensory pathway.

**msq mutation ILK_{L308P} disrupts binding of ILK to β-parvin (Affixin)**

The msq mutation resides in the C terminus of ILK, where, in addition to the kinase domain, several protein-binding domains are located, including those for Paxillin and β-parvin, two proteins known to be expressed in the zebrafish heart (Yamaji et al. 2001). Therefore, in a coimmunoprecipitation assay with [35S]-labeled cotranslated proteins, we evaluated whether the msq mutation ILK_{L308P} disrupts ILK binding to either Paxillin and/or β-parvin. As shown in Figure 5C, both wild-type ILK (hIlk) [lane 4] and mutant hILK_{L308P} [lane 5] bind hPaxillin. In contrast, whereas wild-type hILK is able to bind hβ-parvin (Fig. 5C, lane 1), hILK_{L308P} (Fig. 5C, lane 2) is not. The same result was obtained when [35S]-labeled ILK_{wt} was incubated with purified recombinant GST-Paxillin or GST-β-parvin, respectively (data not shown). To further determine if disruption of ILK–β-parvin binding accounts for the msq mutant heart phenotype, we injected another mutant form of ILK (hIlk_{E359K}) with greatly reduced β-parvin binding (Yamaji et al. 2001) into offspring derived from intercrossing msq heterozygous +/− zebrafish. However, in contrast to hIlk_{wt}, hIlk_{E359K} RNA injection at various concentrations (300–500 pg) did not reconstitute ventricular contractility in msq mutant embryos [0 out of 44] (Fig. 5B), indicating that binding of ILK to β-parvin is essential for maintenance of cardiac ventricular contractility.

**Antisense oligonucleotide mediated knockdown of zebrafish β-parvin (Affixin) phenocopies the msq mutant heart phenotype.**

We demonstrated above that the msq mutation ILK_{L308P} specifically inhibits ILK–β-parvin binding without affecting the interaction with Paxillin, and that the physical interaction of ILK and β-parvin is essential to main-
tain cardiac contractility. The in vivo function of β-parvin has not yet been evaluated in vertebrates.

As shown in Figure 5, I and J, β-parvin and ILK colocalize at sarcomeric Z-discs of rat heart and zebrafish skeletal muscle cells. In msq mutants, β-parvin is also localized at costameres of cardiomyocytes and skeletal muscle cells, indicating that ILK–β-parvin binding is not essential for β-parvin localization within muscle cells [data not shown]. To gain further insight into the role of β-parvin in vivo, we designed a morpholino-modified antisense oligonucleotide directed against the splice donor site of exon 5 [MO-zafx] of zβ-parvin [Affixin] [GenBank accession no. DQ499602] and injected it into wild-type zebrafish embryos at the one-cell stage. When injecting 2.7 ng of MO-zafx into zebrafish wild-type embryos, >60% of the injected embryos [51 out of 86] revealed a phenotype identical to the one observed in msq mutant embryos [Fig. 5D–H; Supplementary Movie 3]. In the MO-zafx-injected embryos, fractional shortening of the ventricle decreased down to 0.57 ± 1.8% by 72 hpf [Fig. 5F]. Similar to msq mutant hearts, the expression of the stretch-responsive factor anf was also found to be severely down-regulated in MO-zafx-injected embryos, in comparison to MO-control-injected embryos [Fig. 5G,H].

Isolation of RNA from MO-zafx-injected embryos confirmed the predicted effect on mRNA splicing, namely, either skipping of exon 5 [129-bp product] or integration of intron 5 [300-bp product], both leading to premature termination of translation. Wild-type zafx cDNA [266-bp product] could not be detected in the injected embryos even at 72 hpf [Fig. 5F, inset]. In contrast to the knockdown of β-parvin, knockdown of Paxillin via injection of morpholino-modified antisense oligonucleotides did not phenocopy the msq mutant phenotype [data not shown], underlining the specific role of β-parvin in the regulation of heart contractility.

siRNA-mediated knockdown of parvins was recently found to result in concomitant decrease in ILK levels, most likely due to protein degradation [Fukuda et al. 2003]. Therefore, the cardiac phenotype of β-parvin-depleted zebrafish embryos might be due to reduced ILK protein levels. Indeed, we also observed severely reduced ILK protein levels in β-parvin-deficient zebrafish embryos, demonstrating that β-parvin is essential in vivo to maintain ILK stability and hence cardiac contractility.

Discussion

The sensing of mechanical stimuli [mechano-sensing] and the transduction of these signals into cellular responses [mechano-transduction] are hallmarks of the vertebrate heart and vital to its function. However, the integrative components that allow cardiac myocytes to sense mechanical strain/stress and adjust their force of contraction are only poorly defined. Here we show that ILK is a novel component of such a cardiac stretch sensor, fulfilling both a structural role as a mechanical integration site that links membrane-bound β-Integrins via β-parvin [Affixin] and α-Actinin to the sarcomeric Z-disc, and a functional role in the regulation of cardiomyocyte contractility via PKB/VEGF signaling.

We describe here a mutation in zebrafish ilk, main squeeze, which impairs cardiac contractility and leads to reduced expression of the stretch-responsive factors ANF and VEGF. ILK specifically localizes to costameres and sarcomeric Z-discs, the mechanical integration sites of cardiomyocytes. msq mutant ILKE359K displays reduced ILK kinase activity. ILK was recently shown to regulate vegf transcription by PKB activation during tumor angiogenesis [Tan et al. 2004]. Similarly, we demonstrate here that both PKB and VEGF act downstream from ILK in the vertebrate heart, since the msq heart failure phenotype can be rescued by ectopic expression of constitutively active PKB or VEGF. Furthermore, the msq mutation specifically disrupts binding of ILK to the Z-disc protein β-parvin, and accordingly cardiac contractility cannot be restored by ectopic expression of β-parvin-binding-deficient ILKE359K. Antisense oligonucleotide-mediated knockdown of zebrafish β-parvin leads to a cardiac phenotype identical to msq. Injected embryos develop heart failure and show severely reduced cardiac expression of the stretch-responsive factor ANF. Therefore, we postulate that the vertebrate heart uses the Integrin–ILK–β-parvin network to sense mechanical stretch and to respond with increased expression of ANF and VEGF, the latter of which is known to augment cardiac force by increasing the heart’s calcium transients [Röttbauer et al. 2005].

Molecular pathways controlling cardiac function in general and the role of ILK in particular are difficult to dissect genetically in placental animals because disturbance of the cardiovascular system leads to early embryonic death. Nullizygous ilk mice die shortly after implantation [Sakai et al. 2003], and mice with endothelial cell-specific deletion of ILK confer placental insufficiency and die at embryonic day 11.5 [E11.5] [Friedrich et al. 2004]. In contrast to mammals, intact cardiovascular function is not essential for the early development of the zebrafish embryo, because it obtains adequate oxygen by diffusion from its environment [Burggren and Pinder 1991; Stainier and Fishman 1994]. Zebrafish msq mutant embryos survive for days to late larval stages, permitting here, for the first time, a detailed analysis of the role of ILK in vertebrate cardiac function.

Using gene knockdown and knockout strategies in mice, proteins of the costameric multiprotein complex as well as of the sarcomeric Z-disc were recently proposed to be involved in mechano-transduction and regulation of cardiac contractility [Samaré 2005]. For instance, ventricular cardiomyocyte-specific disruption of β-Integrin function in mice results in perinatal lethality and, in milder cases, in abnormal cardiac performance with reduced basal contractility and relaxation of the heart [Keller et al. 2001]. Similarly, mice deficient in Melusin, a β1-Integrin-binding protein, develop heart dilatation and contractile dysfunction when subjected to pressure overload by aortic banding [Brancaccio et al. 2003], while overexpression of Melusin protects from dilated cardiomyopathy [De Acetis et al. 2005]. Mice defi-
cipient in the Z-disc protein MLP also develop progressive dilated cardiomyopathy [Hongo et al. 2000]. Interestingly, MLP-deficient heart muscle cells fail to respond to passive stretch by increased contractile force or a compensatory gene program [i.e., anf expression] that aims to augment contractile force. Similar to these mice models with disturbed cardiac stretch sensor function, ILK-deficient msq mutant zebrafish embryos display reduced expression of the stretch-responsive factors anf and vegf in the heart, and consequently develop heart failure. ILK is known to bind to cytosolic β-Integrin subunits and, as shown here, to specifically localize to costameres and Z-discs of cardiomyocytes. This role, connecting transmembrane receptors to sarcomeric structures, makes ILK a likely candidate responsible for Integrin-mediated mechano-sensing/mechano-transduction in the heart. In this context, it is interesting to note that ILK contains four ankyrin domains at its N terminus. Ankyrin repeat domains are found in many proteins that interact with membrane transporters, receptors, and cell adhesion molecules, and therefore supposedly have a role in sensing and responding to mechanical strain. Ankyrin repeat domains recently were found to exhibit tertiary-structure-based elasticity and to behave as nanosprings [Lee et al. 2006]. Thus, ILK might also provide with its ankyrin repeat domains the structural prerequisites for acting as an elastic mechano-sensor in cardiomyocytes. There is also evidence that ILK itself is involved in the sensing of mechanical signals in non-muscle cells. Nho and colleagues have recently shown that matrix-derived mechanical forces sensed by β1-Integrin are capable of modulating ILK activity downstream of PI3K, which in turn regulates fibroblast viability via PKB-dependent mechanisms [Nho et al. 2005].

In some circumstances, ILK has been shown to be important for cell proliferation, differentiation, or cell survival [Guo and Wu 2002; Miller et al. 2003; Bock-Marquette et al. 2004]. Flies and worms deficient in ILK are viable but manifest muscle detachment [Zervas et al. 2001; Mackinnon et al. 2002], and endothelial cell-specific deletion of ILK in mice confers placental insufficiency and leads to increased apoptosis [Friedrich et al. 2004]. However, in msq mutants, proliferation, differentiation, and survival are normal in both cardiomyocytes and skeletal muscle cells, suggesting that its role is more functional than structural in the vertebrate heart.

The msq mutant heart phenotype cannot be suppressed by ectopic expression of kinase-deficient ILK [ILK<sup>Δ11A</sup>], pointing toward an essential role of ILK kinase activity in the pathogenesis of the msq heart failure phenotype. The msq mutation resides within the C-terminal kinase domain of ILK, and ILK kinase activity of msq mutant ILK<sup>Δ308P</sup> is significantly down-regulated. Basal ILK kinase activity is usually low, but is stimulated in a phosphatidylinositol (PI) 3 kinase-dependent manner [Dedhar 2000]. PI3,4,5-triphosphate binding as well as [auto] phosphorylation appear to be crucial for ILK activation, since a mutation in the PH domain [Arg211Ala] renders the ILK kinase inactive [Persad et al. 2001a]. In contrast, the skeletal muscle phenotype of ILK-deficient flies and worms can be rescued by kinase-dead ILK, indicating that kinase activity of ILK is not essential in invertebrates and that the main function of ILK may be a structural adaptor between the plasma membrane and the cytoskeleton at sites of Integrin-mediated cell adhesion [Zervas et al. 2001; Mackinnon et al. 2002].

Many downstream proteins have been identified as in vivo targets of ILK kinase activity, including PKB. Several studies indicate that the kinase activity of ILK is directly responsible for the phosphorylation of Ser 473 and thereby full activation of PKB [Delcommenne et al. 1998; Persad et al. 2000, 2001b, Zhang et al. 2002; Attwell et al. 2003]. However, the mechanism by which ILK promotes PKB phosphorylation is controversial, as other studies suggest that ILK may function in the role of an adaptor protein rather than a kinase in regulating PKB activity [Legate et al. 2006]. We show here that ectopic expression of constitutively active PKB reconstitutes cardiac function in msq mutant embryos, implicating PKB as a downstream effector of ILK in the zebrafish heart.

We identified the msq zebrafish mutant by analyzing cardiac expression of the stretch-responsive factors anf and vegf, both of which we found to be severely down-regulated in msq mutant hearts. Interestingly, ILK–PKB signaling is a known regulator of vegf transcription during tumor angiogenesis [Tan et al. 2004]. Therefore, one might postulate that VEGF expression in msq mutant hearts is reduced due to disturbed ILK–PKB signaling. Accordingly, as demonstrated here, ectopic expression of VEGF can directly reconstitute cardiac contractility in msq mutant embryos. Our findings suggest that the msq mutation in ILK leads to decreased kinase activity, resulting in reduced phosphorylation of PKB and, consequently, in reduced transcription of vegf, which is known to be essential for heart contractility by modulating calcium cycling of cardiomyocytes [Rottbauer et al. 2005]. anf transcription also depends on PKB activation and subsequent phosphorylation/inhibition of GSK-3β [Morisco et al. 2000], both known downstream effectors of ILK. PKB has recently been shown to be activated in response to mechanical forces in a variety of different cell types, including endothelial cells [Dimmeler et al. 1998], vascular smooth muscle cells [Sedding et al. 2005], and heart muscle cells [Petroff et al. 2001].

ILK is known to interact with β-parvin [Affixin], α-parvin [CH-ILKBP], Paxillin, and others via its C terminus [Wu and Dedhar 2001], where the kinase domain and the msq mutation reside. The contractility defects at least in part reflect the interference with β-parvin binding. The ILK<sup>Δ308P</sup> mutation specifically disrupts binding to β-parvin, while interaction with Paxillin is not disturbed. Another ILK mutant, ILK<sup>Δ590K</sup>, with greatly reduced binding to β-parvin [Yamaji et al. 2001] also cannot rescue the msq mutant phenotype. However, since ILK<sup>Δ590K</sup> was also found to be reduced in its kinase activity [Novak et al. 1998], failure to rescue the msq mutant phenotype might be due to both kinase and β-parvin-binding deficiency of this mutant. Further-
more, knockdown of β-parvin leads to the same selective cardiac phenotype as observed in msq mutant embryos, most likely by interference with ILK stability.

In vertebrates, the heart and vasculature have great ability to match function to hemodynamic needs. Some of this is due to homeostatic mechanisms intrinsic to the heart, such as sympathetic nerve activation and adrenal medullary release of catecholamines. In addition, there is a powerful system intrinsic to the heart that matches the input to the output of the heart. This so-called Frank-Starling relationship, first described in 1918, is manifest even in an isolated heart (Starling 1918). The work reported here suggests for the first time that ILK, and associated proteins such as β-parvin, are essential components of a mechanical stretch sensor in the heart, and therefore might play a crucial role in the intrinsic adaptation of the heart to continually changing demands.

Materials and methods

Histology, transmission electron microscopy, immunostaining, and RNA antisense in situ hybridization

For histology, embryos were fixed in 4% paraformaldehyde and embedded in JB-4 (Polysciences, Inc.). Five-micrometer sections were cut, dried, and stained with hematoxylin/eosin. Electron micrographs were obtained essentially as described (Rottbauer et al. 2001). Whole-mount RNA in situ hybridization was carried out essentially as described (Jowett and Lettice 1994) using a digoxigenin-labeled full-length z

Microscopy

In vitro kinase assay

To test for kinase activity of zILK, glutathione-Sepharose-immobilized kinase was incubated with 2 μg of dephosphorylated MBP [Upstate Biotechnology]. Incubations were carried out with 10 μCi of [32P]-γ-ATP at 30°C for 1 h in kinase buffer containing 20 mM HEPES (pH 7.5), 5 mM MgCl2, 1 mM MnCl2, 0.05% NP-40, 7.15 mM NaCl, and ILK E359K, the PKB constructs myrPKB and inactPKB, and pCMV zilk and hilk. RNA was diluted [15, 50, 150 ng/μL in solution A [0.1% phenol red, 0.2 M KCl]] and micro-injected into one-cell-stage embryos. Siblings from the same pool were injected using solution A as control.

Expression and purification of recombinant ILK

The msq and hilk constructs were introduced into hilk by PCR-mutagenesis. hilk was amplified by PCR and subcloned into the pCDNA3.1 vector [Invitrogen]. For expression of zebrafish GST fusion proteins, zilk and hilk were amplified by PCR and subcloned into the pGEX6P-1 vector [Amersham]. GST-zilk fusion proteins were expressed in Escherichia coli BL21 DE3 by inducing bacteria with 1 mM IPTG for 6 h at 16°C. Bacteria were harvested by centrifugation and resuspended in extraction buffer [300 mM NaCl, 50 mM Tris/HCl at pH 7.5, 1 mM DTT, 5 mM EDTA, 5 mM EGTA, 0.01% IgePal, and protease inhibitors Leupeptin, PepstatinA, Aprotinin [10 µg/mL each], and AEBSEF [0.1 mM]]. Bacterial cells were disrupted by sonification, and cell debris was removed by centrifugation [35,000 rpm, 30 min, 4°C]. GST fusion proteins were purified with glutathione-Sepharose matrix [Amersham] as described by the manufacturer.

In vitro translation of proteins and in vitro communoprecipitations

[35S]-Metionine-labeled hILK or hILK was cotranslated with HA-tagged hAfx, hPxn, or hp27 Kip1, respectively, using a TNT-T7 Quick Coupled Transcription/Translation System reaction [Promega]. HA-tagged proteins were immunoprecipitated with HA-antibody HA.11 (Babc0), immobilized to Protein A Sepharose (Sigma). The resin was washed three times with buffer containing 250 mM NaCl, 50 mM Tris, 1 mM EDTA, and 0.01% IgePal (pH 7.5). Bound proteins were eluted by boiling in twofold SDS sample buffer and separated by SDS-PAGE. Proteins were visualized by Coomassie staining, and radiolabeled proteins were detected by autoradiography of the dried gel.

Acknowledgments

We thank S. Marquart and H. Hossler for excellent technical assistance, and A. Doherty for her support in fish care and breeding. We thank S. Dedhar for kindly providing hilk, hilk, and hilk. Chan for supplying us with the PKB constructs.
myrPKB and inactPKB, and S. Yamaji for the anti-β-parvin (Affixin) antibody. M.G. is grateful to U. Fischer for continuous support and helpful discussions. This work was supported by grants from NIH-SRO1HL49579, SRO1DK55383, R01HL63206 (to M.C.F.), DFG Ro2173/1-1, DFG Ro2173/2-1, and BMBF 01GS0420, as well as Klaus-Georg and Sigrid Hengstberger Stipendium [to W.R.], and the Postdoc-Fellowship of the medical faculty of the University of Heidelberg [to I.G.H. and T.D.].

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Integrin-linked kinase, a novel component of the cardiac mechanical stretch sensor, controls contractility in the zebrafish heart


Genes Dev. 2006, 20:
Access the most recent version at doi:10.1101/gad.1448306

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